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Weihua Huang
New York Medical College

Caroline Ojaimi

John T. Fallon
New York Medical College

Dante Travisany

Alejandro Maass

See next page for additional authors

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Authors

Weihua Huang, Caroline Ojaimi, John T. Fallon, Dante Travisany, Alejandro Maass, Larisa Ivanova, Alexandra Tomova, Daniel Gonzalez-Acuna, Henry P. Godfrey, and Felipe C. Cabello

Genome Sequence of *Borrelia chilensis* VA1, a South American Member of the Lyme Borreliosis Group

Weihua Huang,^a Caroline Ojaimi,^a John T. Fallon,^a Dante Travisany,^c Alejandro Maass,^c Larisa Ivanova,^b Alexandra Tomova,^{b,d} Daniel González-Acuña,^e  Henry P. Godfrey,^a Felipe C. Cabello^b

Departments of Pathology^a and Microbiology and Immunology,^b New York Medical College, Valhalla, New York, USA; Center of Mathematical Modeling and Center for Genomic Regulation, Universidad de Chile, Santiago, Chile^c; Faculty of Medicine, Institute of Physiology, Comenius University, Bratislava, Slovakia^d; Faculty of Veterinary Sciences, Universidad de Concepción, Chillán, Chile^e

***Borrelia chilensis* strain VA1 is a recently described South American member of the *Borrelia burgdorferi sensu lato* complex from Chile. Whole-genome sequencing analysis determined its linear chromosome and plasmids lp54 and cp26, confirmed its membership in the Lyme borreliosis group, and will open new research avenues regarding its pathogenic potential.**

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Address correspondence to Henry P. Godfrey, hgodfrey@nymc.edu.

The *Borrelia* genus contains three major groups of species (1, 2): the Lyme borreliosis group, several members of which cause Lyme disease throughout the Northern Hemisphere; the relapsing fever group, the members of which cause relapsing fever worldwide; and the reptile-associated group, the members of which infect reptiles but are not known to cause disease in humans. We recently identified and cultured a new borrelial species from Chile belonging to the Lyme borreliosis group (2). This new species, *Borrelia chilensis* VA1, isolated from *Ixodes stilesi* ticks present on environmental vegetation and long-tailed rice rats, has extended the range of the Lyme borreliosis group *Borrelia* to South America and the Southern Hemisphere.

We took advantage of high-throughput next-generation sequencing for whole-genome analysis, despite our inability to grow *B. chilensis* VA1 free of contaminating *Delftia* species (2). Genomic DNA was isolated from cultured spirochetes using the Qiagen (Valencia, CA) DNeasy blood and tissue kit. Two libraries were separately generated using the Illumina (Hayward, CA) Nextera XT DNA preparation kit and sequenced using a 251-bp paired-end library on the Illumina MiSeq system. The outputs were combined for a total of 13 million paired-end reads, reaching an estimated average coverage of about 50-fold. The sequences were assembled *de novo* using the Velvet algorithm with optimized *k*-mers (3). The resulting high-quality assemblies were mapped to *Borrelia burgdorferi* B31 (4) and *Borrelia garinii* BgVir (5) as reference genomes, and contaminating *Delftia* genomic DNA sequences were removed.

We obtained two complete contigs, one for linear plasmid lp54, and the other for circular plasmid cp26. We also identified 11 contigs for a scaffold of the chromosome. Short gaps in this scaffold were later closed by *in silico* analysis of mapping reads and contigs to genome references. The sequence redundancies between the *de novo* and mapped-to-reference assemblies were identical. The sequence annotations for both the chromosome and plasmids were performed using both Prokka 1.8 (6) and the NCBI

Prokaryotic Genome Annotation Pipeline (7). No clustered regularly interspaced short palindromic repeats (CRISPR) were detected.

The linear chromosome contains 900,694 bp (G+C content, 28.5%), with 812 coding sequences (CDSs), 32 tRNAs, and five rRNAs. The linear plasmid lp54 contains 54,418 bp, with 64 CDSs, and circular plasmid cp26 contains 27,126 bp, with 27 CDSs. Pan-genomic analysis using PGAP-1.11 (8) revealed that components on the chromosome and two plasmids of *B. chilensis* VA1 are syntenic with those of *B. burgdorferi* B31 and *B. garinii* BgVir. A whole-genome comparison with other borrelial species further confirmed *B. chilensis* VA1 as a new genospecies in the Lyme borreliosis group. Additionally, we identified a unique gene, *ndoR*, encoding a naphthalene 1,2-dioxygenase system ferredoxin-NAD⁺ reductase component, the role of which is uncharacterized in *B. chilensis*. Other plasmids containing more repetitive sequences remain to be assembled. The pathogenic potential of this new borrelial species for rodents and humans remains undetermined.

Nucleotide sequence accession numbers. The complete chromosome sequence of *B. chilensis* VA1 and the sequences of lp54 and cp26 have been deposited in the GenBank database under the accession numbers CP009910 to CP009912. This is the first version of the genome sequences for *B. chilensis* VA1.

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REFERENCES

- Margos G, Vollmer SA, Ogden NH, Fish D. 2011. Population genetics, taxonomy, phylogeny and evolution of *Borrelia burgdorferi sensu lato*. *Infect Genet Evol* 11:1545–1563. <http://dx.doi.org/10.1016/j.meegid.2011.07.022>.

2. Ivanova LB, Tomova A, González-Acuña D, Murúa R, Moreno CX, Hernández C, Cabello J, Cabello C, Daniels TJ, Godfrey HP, Cabello FC. 2014. *Borrelia chilensis*, a new member of the *Borrelia burgdorferi sensu lato* complex that extends the range of this genospecies in the Southern Hemisphere. *Environ Microbiol* 16:1069–1080. <http://dx.doi.org/10.1111/1462-2920.12310>.
3. Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
4. Fraser CM, Casjens S, Huang WM, Sutton GG, Clayton R, Lathigra R, White O, Ketchum KA, Dodson R, Hickey EK, Gwinn M, Dougherty B, Tomb JF, Fleischmann RD, Richardson D, Peterson J, Kerlavage AR, Quackenbush J, Salzberg S, Hanson M, van Vugt R, Palmer N, Adams MD, Gocayne J, Weidman J, Utterback T, Wattley L, McDonald L, Artiach P, Bowman C, Garland S, Fuji C, Cotton MD, Horst K, Roberts K, Hatch B, Smith HO, Venter JC. 1997. Genomic sequence of a Lyme disease spirochaete, *Borrelia burgdorferi*. *Nature* 390:580–586. <http://dx.doi.org/10.1038/37551>.
5. Brenner EV, Kurilshikov AM, Stronin OV, Fomenko NV. 2012. Whole-genome sequencing of *Borrelia garinii* BgVir, isolated from taiga ticks (*Ixodes persulcatus*). *J Bacteriol* 194:5713. <http://dx.doi.org/10.1128/JB.01360-12>.
6. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.
7. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufo S, Li W. 2013. Prokaryotic genome annotation pipeline. The NCBI Handbook, 2nd ed. National Center for Biotechnology Information, Bethesda, MD.
8. Zhao Y, Wu J, Yang J, Sun S, Xiao J, Yu J. 2012. PGAP: pan-genomes analysis pipeline. *Bioinformatics* 28:416–418. <http://dx.doi.org/10.1093/bioinformatics/btr655>.