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Durga S. Borkar
Nisha R. Acharya
Chelsia Leong
Prajna Lalitha
Muthiah Srinivasan

See next page for additional authors

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Authors
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Cytotoxic clinical isolates of *Pseudomonas aeruginosa* identified during the Steroids for Corneal Ulcers Trial show elevated resistance to fluoroquinolones

Durga S Borkar¹, Nisha R Acharya¹,²,⁵, Chelsia Leong³, Prajna Lalitha⁴, Muthiah Srinivasan⁴, Catherine E Oldenburg², Vicky Cevallos², Thomas M Lietman¹,²,⁵, David J Evans³,⁶ and Suzanne M J Fleiszig³,⁷*

**Abstract**

**Background:** To determine the relationship between type three secretion genotype and fluoroquinolone resistance for *P. aeruginosa* strains isolated from microbial keratitis during the Steroids for Corneal Ulcers Trial (SCUT) and for two laboratory strains, PA103 and PAO1.

**Methods:** Confirmed *P. aeruginosa* isolates from the SCUT were divided into exoU(+) or exoU(−). The exoU(+) strains contained the gene encoding ExoU, a powerful phospholipase toxin delivered into host cells by the type three secretion system. Isolates were then assessed for susceptibility to fluoroquinolone, cephalosporin, and aminoglycoside antibiotics using disk diffusion assays. Etest was used to determine the MIC of moxifloxacin and other fluoroquinolones. Laboratory isolates in which the exoU gene was added or deleted were also tested.

**Results:** A significantly higher proportion of exoU(+) strains were resistant to ciprofloxacin (p = 0.001), gatifloxacin (p = 0.003), and ofloxacin (p = 0.002) compared to exoU(−) isolates. There was no significant difference between exoU(+) or exoU(−) negative isolates with respect to susceptibility to other antibiotics except gentamicin. Infections involving resistant exoU(+) strains trended towards worse clinical outcome. Deletion or acquisition of exoU in laboratory isolates did not affect fluoroquinolone susceptibility.

**Conclusions:** Fluoroquinolone susceptibility of *P. aeruginosa* isolated from the SCUT is consistent with previous studies showing elevated resistance involving exoU encoding (cytotoxic) strains, and suggest worse clinical outcome from infections involving resistant isolates. Determination of exoU expression in clinical isolates of *P. aeruginosa* may be helpful in directing clinical management of patients with microbial keratitis.

**Keywords:** *P. aeruginosa*, Microbial keratitis, SCUT, Fluoroquinolone resistance, ExoU
contact lens-associated cases [20]. Interestingly, several other studies have also shown a relationship between exoU expression and resistance of P. aeruginosa to contact lens disinfectants [21], and to multiple antimicrobials [22], especially fluoroquinolones [23]. In the latter instance, it was shown that P. aeruginosa clinical isolates encoding exoU were more likely to encode multiple mutations in quinolone target genes, e.g. gyrA and parC, and suggested a co-evolution of these traits [24].

The Steroids for Corneal Ulcers Trial (SCUT), was a randomized controlled trial investigating the effects of adjunctive corticosteroid therapy to fluoroquinolone (moxifloxacin) treatment on the outcomes of bacterial keratitis [25,26]. Analysis of moxifloxacin resistance of bacterial keratitis isolates (including P. aeruginosa) from the SCUT study showed that increased resistance to this fluoroquinolone in vitro (i.e. increased minimum inhibitory concentration) was associated with poorer clinical outcomes, including reduced visual acuity at 3 weeks, a larger ulcer, and slower corneal re-epithelialization [27]. Here, we analyzed P. aeruginosa strains isolated from the SCUT study, and two laboratory isolates and their T3SS mutants, for their in vitro susceptibility to fluoroquinolones and other antimicrobials including cephalosporins and aminoglycosides. Antimicrobial classes and individual agents tested were chosen because of their common clinical empirical use against bacterial keratitis [28]. The objective was to determine if the SCUT clinical isolates and laboratory strains of P. aeruginosa also showed a relationship between fluoroquinolone resistance and exoU expression.

Methods

a) Clinical and laboratory isolates of P. aeruginosa and T3SS mutants

The P. aeruginosa isolates used were obtained from the Steroids for Corneal Ulcers Trial (SCUT), a randomized controlled trial investigating the effect of adjunctive corticosteroids on outcomes in bacterial keratitis [25]. The SCUT study was conducted with the approval of the University of California, San Francisco; Aravind Eye Hospital, Madurai, India; Dartmouth-Hitchcock Medical Center, Hanover, NH. Informed consent was obtained from all study participants. Corneal isolates were collected from all study participants. Corneal isolates were collected from patients with culture-confirmed bacterial keratitis. The isolates used in the present study were previously confirmed as P. aeruginosa using growth morphology, Gram stain, and API test strips, and their type three secretion gene profile was determined [29]. Presence of exoU was determined by polymerase chain reaction using primers and conditions described in detail by Ledbetter et al. [30]. Two wild-type laboratory strains of P. aeruginosa (PA103, encodes exoU) and PAO1 (does not encode exoU) were tested. For strain PA103, two T3SS mutants PA103exsA+Ω and PA103ΔexoU were also used. Strain PAO1 was complemented with a plasmid encoding exoU (pUCPexoU) or a plasmid control (pUCP18).

b) Antibiotic susceptibility testing

Susceptibility to moxifloxacin, the fluoroquinolone antibiotic used in the SCUT, was measured using the Etest method (AB BIODISK, Solna, Sweden) for all isolates using a standardized inoculum of 1 × 10⁸ CFU/mL on Mueller-Hinton agar plates, which were then incubated at 35°C. Disc diffusion susceptibility testing was performed for various other antibiotics (other fluoroquinolones, cephalosporins, and aminoglycosides) and P. aeruginosa strains classified as susceptible, intermediate, or resistant to each antibiotic tested based on the Clinical and Laboratory Standards Institute (CLSI) guidelines [31]. Since CLSI guidelines do not provide susceptibility ranges for the MIC of moxifloxacin for P. aeruginosa, susceptibility ranges for ciprofloxacin were used for Etest results. Laboratory personnel were masked to the clinical data.

c) Visual acuity

SCUT patients with culture-confirmed bacterial keratitis were evaluated at multiple time points, including enrollment and three months, by certified refractionists who performed visual acuity examinations. Visual acuity was measured as best spectacle-corrected visual acuity (BSCVA) in logMAR units, with 0.1 logMAR corresponding to approximately one line of acuity. Laboratory personnel were masked to the clinical data.

d) Statistical analysis

Differences in disc diffusion susceptibility for various antibiotics, and mean minimum inhibitory concentration (MIC) of moxifloxacin, between exoU(+) or exoU(−) strain types were compared using Fisher’s exact test and the Student’s t-Test, respectively. The t-Test was also used to compare the change in BSCVA between enrollment and three months between patients with corneal ulcers caused by resistant or non-resistant exoU(+) P. aeruginosa.

Results

Of 500 patients enrolled in the SCUT, 110 had a corneal ulcer involving P. aeruginosa [32]. Confirmed P. aeruginosa isolates that were available for study were divided into two groups according to presence or absence of the exoU gene. Table 1 shows disc diffusion antimicrobial
susceptibility data for 19 \textit{exoU}(+) and 75 \textit{exoU} (−) isolates. A significantly higher proportion of \textit{exoU}(+) strains were resistant to ciprofloxacin (p = 0.001), gatifloxacin (p = 0.003), and ofloxacin (p = 0.002) compared to \textit{exoU}(−) isolates. There was no significant difference between \textit{exoU}(+) or \textit{exoU}(−) negative isolates with respect to susceptibility to the cephalosporins tested, or to the aminoglycosides amikacin and tobramycin. Interestingly, there were significantly more gentamicin resistant \textit{exoU}(+) isolates (p = 0.02).

Of the 19 \textit{exoU}(+) corneal isolates, 4 were resistant to gatifloxacin, ciprofloxacin, and ofloxacin based on disc diffusion susceptibility testing, and had an MIC greater than or equal to 4 micrograms/ml. The BSCVA of patients with ulcers caused by the resistant ExoU(+) isolates improved, on average, approximately six lines less from enrollment compared to patients with ulcers caused by non-resistant ExoU(+) isolates (−0.38 logMAR vs. -1.04 logMAR) although this difference was not statistically significant (p = 0.08).

In another set of experiments, the MIC of moxifloxacin was determined and compared for 21 \textit{exoU}(+) and 76 \textit{exoU} (−) SCUT isolates of \textit{P. aeruginosa}. Figure 1 shows that isolates encoding \textit{exoU} had, on average, a greater than two-fold higher MIC compared to \textit{exoU} (−) strains (p = 0.0001).

Laboratory isolates of \textit{P. aeruginosa} were also tested to determine if genetic deletion of the T3SS, or deletion or acquisition of \textit{exoU}, affected bacterial susceptibility to fluoroquinolones or other antibiotics using disc diffusion and Etest assays (Table 2). Neither deletion of the T3SS or \textit{exoU} in the cytotoxic strain PA103, nor acquisition of \textit{exoU} in the invasive strain PAO1, affected sensitivity to any antimicrobial tested.

### Table 1 Antibiotic susceptibility\(^1\) of SCUT Isolates of \textit{P. aeruginosa} and T3SS genotype

<table>
<thead>
<tr>
<th>Antibiotic</th>
<th>\textit{exoU}(+) (n = 19)</th>
<th>\textit{exoU} (−) (n = 75)</th>
<th>\text{P-value}(^2)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Fluoroquinolones</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ciprofloxacin(^3)</td>
<td>5 (28%)</td>
<td>13 (72%)</td>
<td>0.001</td>
</tr>
<tr>
<td>Gatifloxacin</td>
<td>5 (26%)</td>
<td>14 (74%)</td>
<td>0.003</td>
</tr>
<tr>
<td>Ofloxacin</td>
<td>6 (32%)</td>
<td>13 (68%)</td>
<td>0.002</td>
</tr>
<tr>
<td><strong>Cephalosporins</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cefotaxime</td>
<td>3 (16%)</td>
<td>16 (84%)</td>
<td>0.70</td>
</tr>
<tr>
<td>Ceftazidime</td>
<td>3 (16%)</td>
<td>16 (84%)</td>
<td>0.20</td>
</tr>
<tr>
<td><strong>Aminoglycosides</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Amikacin</td>
<td>1 (5%)</td>
<td>18 (95%)</td>
<td>0.37</td>
</tr>
<tr>
<td>Gentamicin</td>
<td>5 (26%)</td>
<td>14 (74%)</td>
<td>0.02</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>3 (16%)</td>
<td>16 (84%)</td>
<td>0.054</td>
</tr>
</tbody>
</table>

\(^1\)Disc diffusion assay.
\(^2\)Fisher’s exact test.
\(^3\)Disc diffusion susceptibility data for ciprofloxacin not available for one \textit{exoU}(+) isolate (n = 18).

### Discussion

Topical fluoroquinolones are commonly used in the treatment of bacterial keratitis, in which \textit{P. aeruginosa} is a leading causative pathogen. The data presented in this study show the antimicrobial resistance patterns of \textit{P. aeruginosa} strains (94) isolated from patients during the
The data show a significant increase in the number of isolates resistant to fluoroquinolones if the bacteria encode the type three secreted cytotoxin ExoU. The data also confirm that deletion or acquisition of exoU gene does not affect fluoroquinolone susceptibility. These data are consistent with previous studies showing an association of exoU expression with fluoroquinolone resistance in *P. aeruginosa*, and support the hypothesis that T3SS expression and fluoroquinolone resistance are co-selected traits in both ocular and non-ocular clinical isolates [22,23]. Indeed, prior studies have found a significantly higher rate of gyrA mutations, combined with mutations in other fluoroquinolone target genes e.g. parC, in exoU encoding *P. aeruginosa* isolates [23,24].

It was of interest that we also found significantly more gentamicin resistant exoU(+) isolates in the study, although this did not occur for other aminoglycosides. This finding is consistent with a previous study looking at bacteremia isolates of *P. aeruginosa* in which the presence of exoU was also associated with increased resistance to gentamicin, but not amikacin [11]. It would be of interest to determine if gentamicin resistance in these ocular exoU(+) isolates relates to specific mutations in drug target genes, as shown for fluoroquinolones [24].

In bacterial keratitis, antibiotic susceptibility likely plays a role in clinical outcome, suggesting a role for testing antibiotic susceptibility in management decisions [27,33]. Interestingly, we found that SCUT patients with ulcers caused by fluoroquinolone resistant exoU(+) *P. aeruginosa* isolates trended toward worse visual outcomes at three months compared to those with ulcers caused by non-resistant exoU(+) isolates. These results could reflect increased bacterial persistence in the cornea, allowing for greater tissue damage from bacterial toxins (including ExoU), and from host immune infiltrative responses.

The continuing emergence of antimicrobial resistant strains has the potential to influence “optimal” treatment of *P. aeruginosa* keratitis. However, *P. aeruginosa* resistance rates to fluoroquinolones and aminoglycosides, common choices for management of this condition, remain relatively low and stable in many places, although they can vary widely by geographical location and time [22,34-36]. Our study, and those of others [11,22-24], however, suggest that the combination of exoU and antibiotic resistance may be particularly deleterious to the patient and rapid identification of these isolates, if possible, could help optimize treatment.

**Conclusion**

Fluoroquinolone susceptibility of *P. aeruginosa* isolated from the SCUT is consistent with previous studies showing elevated resistance involving exoU encoding (cytotoxic) strains, and suggest worse clinical outcome from infections involving these resistant isolates. Increased gentamicin resistance is also associated with exoU(+) strains. Determination of exoU expression in clinical isolates of *P. aeruginosa*, and antibiotic susceptibility testing, may be helpful in directing optimal clinical management of patients with microbial keratitis if methods could be developed with sufficient speed and accuracy, and which could be applied in a clinical setting.

**Competing interests**

Dr. Suzanne Fleiszig is a paid consultant for Allergan, Irvine, CA. That work is unrelated to the content of this manuscript. There are no competing interests for any of the authors.

**Authors’ contributions**

Study design and analysis: DB, SF, NA, DE, PL, MS, TL. Experiments: DB, CL, CO, VC. Manuscript writing: DB, DE, NA, SF. Supervision: SF, NA, TL. All authors read and approved the final manuscript.

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Author details
1Department of Ophthalmology, University of California, 94143 San Francisco, CA, USA. 2Francis I. Proctor Foundation, University of California, 94143 San Francisco, CA, USA. 3School of Optometry, University of California, 688 Minor Hall, 94720 Berkeley, CA, USA. 4Aravind Eye Care System, Madurai, Tamil Nadu, India. 5Department of Epidemiology and Biostatistics, University of California, 94143 San Francisco, CA, USA. 6College of Pharmacy, Touro University California, 94592 Vallejo, CA, USA. 7Programs in Vision Science, Infectious Diseases and Immunity, and Microbiology, University of California, 94720 Berkeley, CA, USA.

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